

Helminth Parasites of Goats Slaughtered at Three Abattoirs in Calabar, Cross River State, Nigeria

Cletus I. Iboh

Cross River University of Technology, Department of Animal and Environmental Biology, Faculty of Biological Sciences,
Calabar, Cross River State, Nigeria

Abstract:- The aim of this investigation was to determine the prevalence rate of endoparasite helminthes in two breeds of goats, West Africa Dwarf (WAD) and Red Sokoto (RS), in the study area. Out of 532 faecal samples examined from slaughtered goats in three abattoirs (Atakpa, Anantigha, and Ikot Eno Obong) in Calabar, 413 (77.6%) were infected with helminth parasites. Red Sokoto goats were more infected (78.35%) than West Africa Dwarf goats (73.2%). The total mean faecal egg counts was highest in Red Sokoto goats 163 ± 10.72 , compared to 146.7 ± 11.38 recorded in WAD goats. The three helminthes recovered from 413 faecal samples of goats were *Haemonchus contortus*, *Trichuris trichiura* and *Taenia* spp. *H. contortus* had the highest prevalence rate of 49.4%, compared to *T. trichiura* 20.3% and *Taenia* spp 7.9%. Female goats were more infected (82.6%) as against 77.9% in males. Female goats had higher total faecal egg counts of 155.2 ± 3.33 than 127.2 ± 3.01 in male goats. It was found that adult goats were more infected (89.6%) than young ones (75.5%). Months and season have influence on the infection rate of parasites on goats. *H. contortus* prevalence was highest (26.1%) in September, while seasonal prevalence was highest (42.1%) in late rainy season. In this study, since more RS goats were infected than WAD goats, it was recommended that small ruminant goat farmers should concentrate more in the farming of WAD goats because of their resilience to parasitic infection than RS goats, in the study area.

Keywords: Helminths parasites, prevalence, WAD and RS goats, Calabar, Cross River.

I. INTRODUCTION

A mong ruminants, cattle, sheep, goats, buffalo, deer, elk, Ageraffes and carmels, sheep and goats are the smallest and are reared in Nigeria for meat, Milk, skin and sales [1-5]. According to FAO there were about 1,050 million goats worldwide in 2014 with about 300 species known to exist [6]. Africa accounts for about 33.1 % and Nigeria with 49 117 654 heads represents about 4.7% of the world output [7]. Goats have the ability to withstand harsh climatic conditions and contrary to sheep that are grazers, goats are browsers that feed mostly on shrubs and tree leaves in adverse environments with low fertility lands unfit for growing crops [8]. Domestic goat is among the earliest animal domesticated by man and is distributed worldwide with higher concentration in tropical areas and dry zones [9-11]. Goats have been found to contribute significantly to the livelihood of most low income earners in Nigeria, mainly for the provision of animal protein and income [12]. Socio-economic importance is attached to

goat such that, in some instances they may be the only realizable wealth of a rural household [13]. Goats are important source of protein for both the rich and poor and a food delicacy highly appreciated during ceremonies and festivals, most especially in the southwestern zone of the country [8]. Goats are excellent meat producers for human consumption in view of its short generation intervals and the absence of religious taboos associated with their meat as they are rich sources of protein and can help bridge the gap of protein malnutrition among consumers [14]. The intestine of goat has been used in making catgut which is still in use as a material for internal human surgical sutures and string for musical instruments [11]. The different indigenous breeds of goats in Nigeria, acclimatized to different ecological zones are the West African Dwarf (WAD), West African long-legged goat and the Red Sokoto [4]. The West African Dwarf goats are more predominant in the south while the Red Sokoto is found in the north [15]. The livestock industry plays a vital role in the economy of Nigeria. It serves as a major source of income and livelihood for majority of Nigerians who are rural settlers and contributes about 5.2% of the National Gross Domestic Product (GDP) [16]. Despite these benefits, helminth infections still cause serious economic losses in Nigeria as a result of reductions in milk production, weight gain, fertility and carcass quality. Ruminants are parasitized by two phyla of helminthes, namely nemathelminthes and platyhelminthes. Nemathelminthes are the round worms which include *Haemonchus*, *Bonostomum*, *Oesophagostomum* and *Charbetia*. Plathelminthes include cestodes (e.g. *Avitellina*, *Moniezia*, *Stilesia* and *Taenia*) and trematodes such as *Dicrocoelium*, *Eurytrema*, *Fasciola* and *Paramphistomum* [17]. Gastrointestinal nematode infections (GIN) are the main prevalent parasitic diseases affecting small ruminant productivity worldwide, especially in tropics and sub-tropics [18-19]. Globally the most common nematode species known to affect small ruminants are *Haemonchus contortus*, *Trichostrongylus colubriformis*, *Teladorsagia circumcincta* and some species such as *Nematodirus* spp, which are not found in sub-Saharan Africa [20]. From the public health point of view, reports of zoonotic meta-cestodes; *Cysticercus bovis* and hydatid cyst [21-23], nematode; *Oesophagostomum* [24-26], and trematodes; *Dicrocoelium dendriticum*, *Eurytrema pancreaticum* and *Fasciola gigantica* [27-28], entering the food chain in Nigeria are of great public

health concern. Human infections with these parasites may result in diarrhoea, retarded growth, intellectual and cognitive retardation [29], cystic echinococcosis and cysticercosis [30]. Goat meat is a delicacy in Calabar city and its environs, where goat-pepper-soup joints are crowded by teenagers and adults in the evening due to the rejection of red meat from cow. The aim of this study was to provide epidemiological information on goat's endoparasitism, which will help in instituting sustainable control programmes against these parasites for consumers in the study area.

II. MATERIALS AND METHODS

2.1 Study Area

This study was carried out in Calabar, South Eastern Nigeria. Calabar is the Capital of Cross River State, whose name was derived from the Cross River which passes through the state. Calabar comprised of two local government areas, Calabar South and Calabar Municipality. This area lies on coordinates 40° 57' 0" N 80° 19' 30" E. The study area has two main seasons; the rainy and dry seasons. The rainy season usually starts in April and ends in October, while the dry season starts in November and ends in March. The Local Government has a sub-equatorial climate and a moderately hot temperature (with an average temperature of 28°C) which does not fluctuate greatly. It is characterized by frequent and high rainfall as well as high relative humidity. The three abattoirs used for this investigation were Atakpa, Anantigha (in Calabar South) and Ikot Eno Obong (in Calabar Municipality). This study covered six month (July to December, 2019). For the purpose of seasonal analysis of result, the months were divided into Early rainy season (July to August), Late rainy season (September to October), and Early dry season (November to December).

2.2 Sample collection and examination

Goats are slaughtered in the earlier mentioned abattoirs between 6.30 a.m. - 8.30 a.m. The ages of the goats were determined by estimation of the teeth [31], and were grouped into two categories, young (≤ 2 year having two permanent teeth) and (≥ 2 years with more than two permanent teeth). A total of 532 faecal samples were collected, 149 from West African Dwarf (WAD) goats and 383 from Red Sokoto (RS) goats in the three aforementioned abattoirs. After killing the goats and removal of abdominal viscera, faeces were collected from the gastrointestinal tract of each goat. Faecal samples were placed into plastic universal vials before transportation to Biological Science laboratory of Cross River University of Technology, Calabar, for parasitological investigation. The concentration method was used in the study of gastrointestinal helminth parasites [32]. The advantage of this method is that it offers high concentration of parasites and is relatively free of contaminating particulate materials. The faecal egg count

per gram (EPG) was determined by the modified McMaster technique, using saturated saline solution to determine the parasitic load [33]. Observed helminth ova and adult were identified using known morphological characteristics [34], before multiplication by a dilution factor of 24 to convert the eggs to egg per gram faeces.

2.3 Data analysis

Data obtained for egg and adult counts were used to show the mean burden of parasites as well as construction of tables. Chi-square analysis was further used to establish the association between parasite egg output and seasons, age and sex.

III. RESULTS

Table 1 shows prevalence of gastrointestinal helminth parasites and mean egg count per gram of faeces among breed of goats. Of the 532 goat faecal samples examined from Atakpa, Anantigha and Ikot Eno Obong slaughter abattoirs, 413 (77.6%) were infected with helminthes parasites, comprising of 109 (73.2%) and 304 (78.4%) of West Africa Dwarf (WAD) and Red Sokoto (RS) goats respectively. Three helminth parasites isolated from WAD and RS goats were 43.6 and 51.7% for *Haemonchus contortus*, 17.4% and 21.4% for *Trichuris trichiura* and 12.1% and 6.3% for *Taenia* species respectively (Table 1). Red Sokoto goats had a higher prevalence rate of 78.35% endoparasitic helminthes compared to 73.2% in West Africa Dwarf goats. There was statistical significant difference ($\chi^2 = 6.3$, $df = 2$, $p < 0.05$) in the prevalence of helminth parasites between breed of goats. Higher worm burden per animal was recorded in *H. contortus* 213.0 \pm 15.89 in RS goats, but lower 180.8 \pm 16.66 in WAD goats. The mean egg count per gram of faeces of *T. trichiura* was higher 164.8 \pm 16.1 in RS goats, but lower 118.0 \pm 12.59 in WAD goats. With respect to *Taenia* spp., the mean faecal egg count was higher 135.3 \pm 11.87 in WAD goats but lower 109.6 \pm 17.4 in RS goats. However, the total mean faecal egg count was higher 163.7 \pm 10.72 in RS goats compared to 146.7 \pm 11.38 recorded in WAD goats. Table 2 illustrates the mean faecal egg count per gram of faeces according to sex of goat. In female goats, the mean egg counts were highest for *H. contortus* (272.0 \pm 17.3) and lowest for *Taenia* spp (56.0 \pm 9.07). For male goats, the mean egg counts were also highest for *H. contortus* (100.8 \pm 9.99) and lowest for *Taenia* spp (46.4 \pm 5.96). *H. contortus* had a higher prevalence for both sexes (53.3% for males and 61.6% for females). *T. trichiura* and *Taenia* spp have a higher prevalence for males (17.3% and 7.4%) respectively, but lower for females (15.2% and 5.1%) respectively. Female goats had a higher total egg count 155.2 \pm 3.33 than 127.2 \pm 3.01 in male goats. There was statistical significant difference ($\chi^2 = 9.3$, $df = 2$, $p < 0.01$) in the prevalence of intensity of eggs per gram of faeces in both sexes. (Table 2).

Table 1: Prevalence of GI helminth parasites and mean egg count per gram of faeces among breed of goats in Calabar abattoirs

| Helminth eggs recovered | Prevalence (%) | EPG | Prevalence (%) | EPG | Total Prevalence (%) |
|-----------------------------|--------------------------------|-------------|-------------------------|-------------|----------------------|
| | West Africa Goat (WAD) n = 149 | Mean±SEM | Red Sokoto (RS) n = 383 | Mean±SEM | WAD + RS n = 532 |
| <i>Haemonchus contortus</i> | 65 (43.6) | 180.8±16.66 | 198 (51.7) | 213.0±15.89 | 263 (49.4) |
| <i>Trichuris trichiura</i> | 26 (17.4) | 118.0±12.59 | 82 (21.4) | 164.8±16.1 | 108 (20.3) |
| <i>Taenia spp</i> | 18 (12.1) | 135.3±11.87 | 24 (6.3) | 109.6±17.4 | 42 (7.9) |
| Total | 109 (73.2) | 146.7±11.38 | 304 (78.35) | 163.7±10.72 | 413 (77.6) |

Table 2: Prevalence of helminth parasites and mean egg count per gram of faeces according to the sex of goat

| Helminth eggs recovered | Prevalence (%) Male (n = 394) | EPG Mean±SEM | Prevalence (%) Female (n = 138) | EPG Mean±SEM |
|-----------------------------|-------------------------------|--------------|---------------------------------|--------------|
| <i>Haemonchus contortus</i> | 210 (53.3) | 100.8±9.99 | 85 (61.6) | 272.0±17.33 |
| <i>Trichuris trichiura</i> | 68 (17.4) | 216.0±11.08 | 21 (15.2) | 135.5±10.91 |
| <i>Taenia spp</i> | 29 (7.4) | 46.4±5.96 | 8 (5.1) | 56.0±9.07 |
| Total | 307 (77.9) | 127.2±3.01 | 114 (82.6) | 155.2±3.33 |

In age related prevalence of eggs in faeces, adult goats have a total higher prevalence of 89.6% than young goats with 75.5%. The adult mean egg counts of *H. contortus* are still highest in adult and young goats (66.9±9.81 and 39.4±8.04) respectively. Adult goats have significantly higher prevalence

rate of 59.0%, 20.6% and 9.2% eggs in their faeces than 53.2%, 16.7%, and 5.6% in young goats' faeces. However, the mean egg counts for *T. trichiura* and *Taenia spp* are higher in adult goats 46.2±5.65 and 60.8±7.36 respectively, than in young goats having 44.3±7.89 and 51.2±3.99 respectively.

Table 3: Prevalence of GI helminth parasites and mean egg counts per gram of faeces according to age of goats slaughtered in Calabar abattoirs

| Helminth eggs recovered | Prevalence (%) Young goats n = 216 | EPG Mean±SEM | Prevalence (%) Adult goats N = 316 | EPG Mean±SEM |
|-----------------------------|------------------------------------|--------------|------------------------------------|--------------|
| <i>Haemonchus contortus</i> | 115 (53.2) | 39.4±8.04 | 189 (59.0) | 66.9±9.81 |
| <i>Trichuris trichiura</i> | 36 (16.7) | 44.3±7.89 | 65 (20.6) | 46.2±5.65 |
| <i>Taenia spp</i> | 12 (5.6) | 51.2±3.99 | 29 (9.2) | 60.8±7.36 |
| Total | 163 (75.5) | 52.3±6.01 | 283 (89.6) | 62.8±4.84 |

Table 4 illustrates the monthly and seasonal recovered adult helminth parasites in goat faeces slaughtered in Calabar abattoirs. A total of 2232 (16.2), 8688 (62.9) and 2880 (20.9) egg per gram of goat faeces were recovered for *Taenia spp*, *H. contortus* and *T. trichiura* respectively. The prevalence of adult *H. contortus* was highest (62.9%) compared with those of *T. trichiura* (20.9%) and *Taenia spp* (16.2%). Equally,

adult *H. contortus* prevalence was highest (26.1%) in September during the late rainy season (Table 4). Figure 1 shows the mean prevalence of adult helminth parasites recovered from faeces of slaughtered goats in Calabar abattoirs. Images of adult *H. contortus*, *Taenia spp* and egg of *T. trichiura* recovered from goat faeces are shown in Plates 1, 3, and 3 respectively.

Table 4: Total monthly and seasonal prevalence of adult helminthes recovered in goats faeces slaughtered in Calabar abattoirs

| Season | Month | <i>Taenia spp</i> EPG | Prevalence (%) | <i>Haemonchus contortus</i> EPG | Prevalence (%) | <i>Trichuris trichiura</i> EPG | Prevalence (%) | Total EPG and prevalence (%) |
|--------------------|-----------|-----------------------|----------------|---------------------------------|----------------|--------------------------------|----------------|------------------------------|
| Early Rainy Season | July | 384 | 17.2 | 1728 | 19.9 | 192 | 6.7 | 2304 (16.7) |
| | August | 288 | 12.9 | 2088 | 24.0 | 336 | 11.7 | 2712 (19.7) |
| Late Rainy Season | September | 768 | 34.4 | 2304 | 26.5 | 528 | 18.3 | 3600 (26.1) |
| | October | 432 | 19.4 | 1296 | 14.9 | 360 | 12.5 | 2088 (15.1) |
| Dry Season | November | 216 | 9.7 | 864 | 9.9 | 576 | 20.0 | 1656 (12.0) |
| | December | 144 | 6.5 | 408 | 4.7 | 888 | 30.8 | 1440 (10.4) |
| Total | | 2232 | 16.2 | 8688 | 62.9 | 2880 | 20.9 | 13800 (100) |

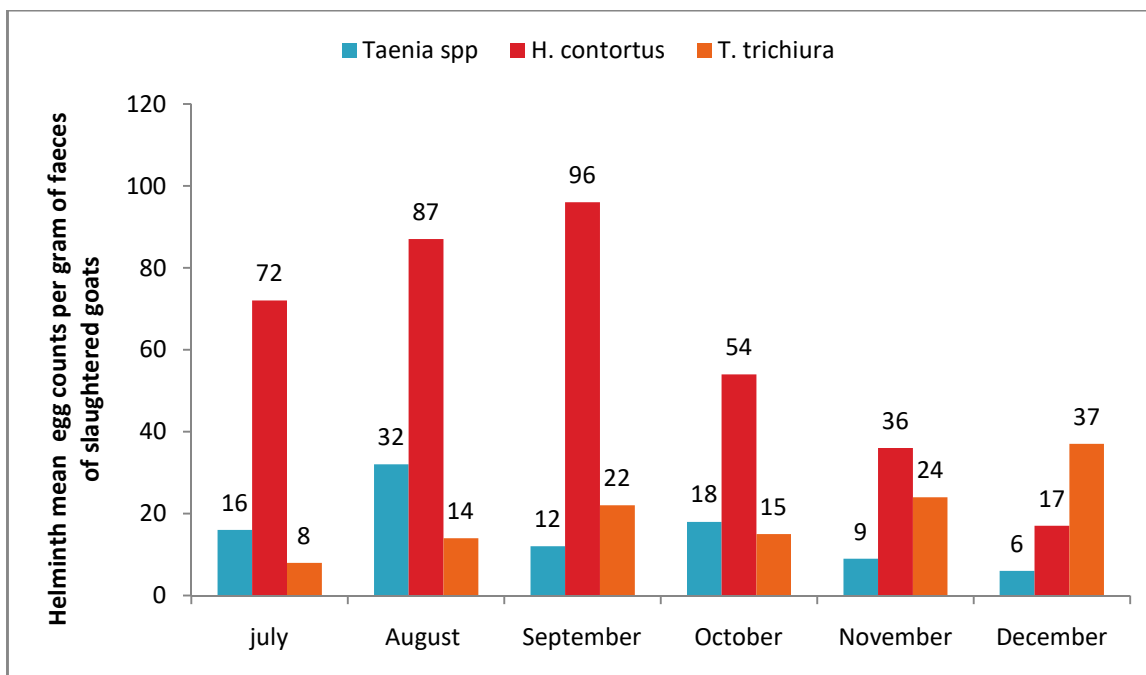


Fig.1: Monthly mean egg counts of helminth parasites per gram of faeces recovered.



Plate 1: Image of adult *Haemonchus contortus* recovered from goat faeces

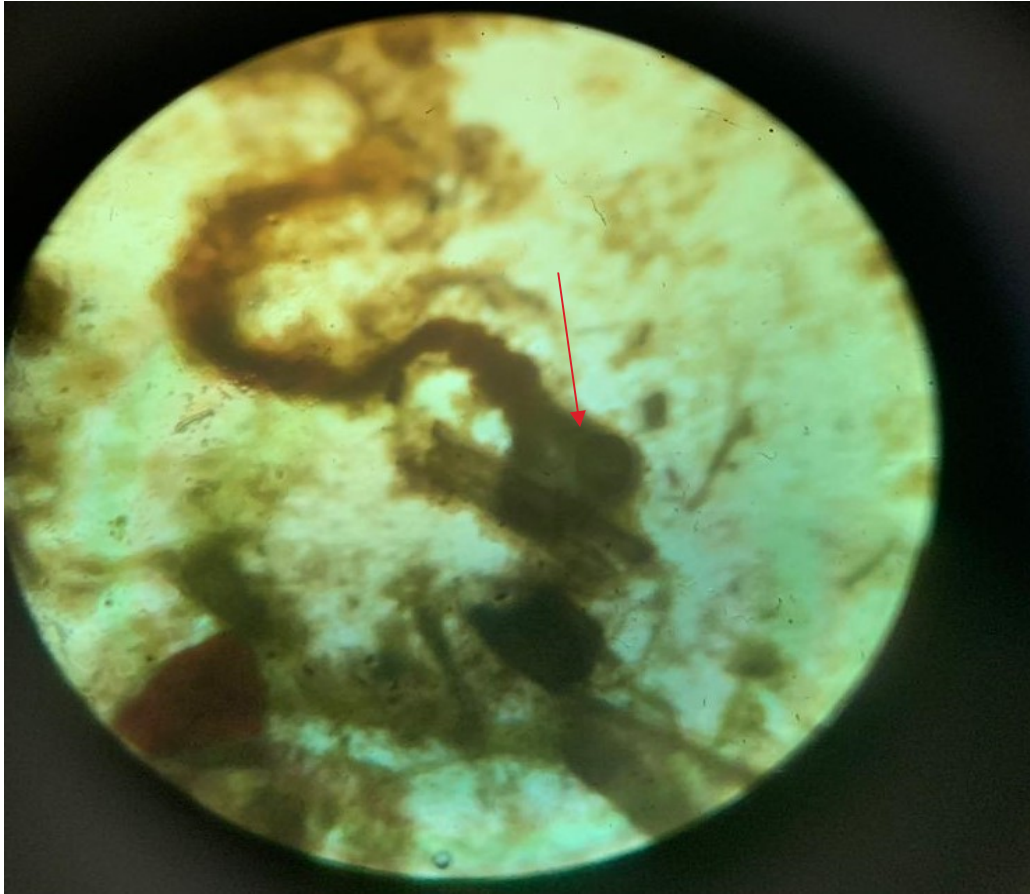


Plate 2: Image of adult *Taenia* spp recovered from goat faeces

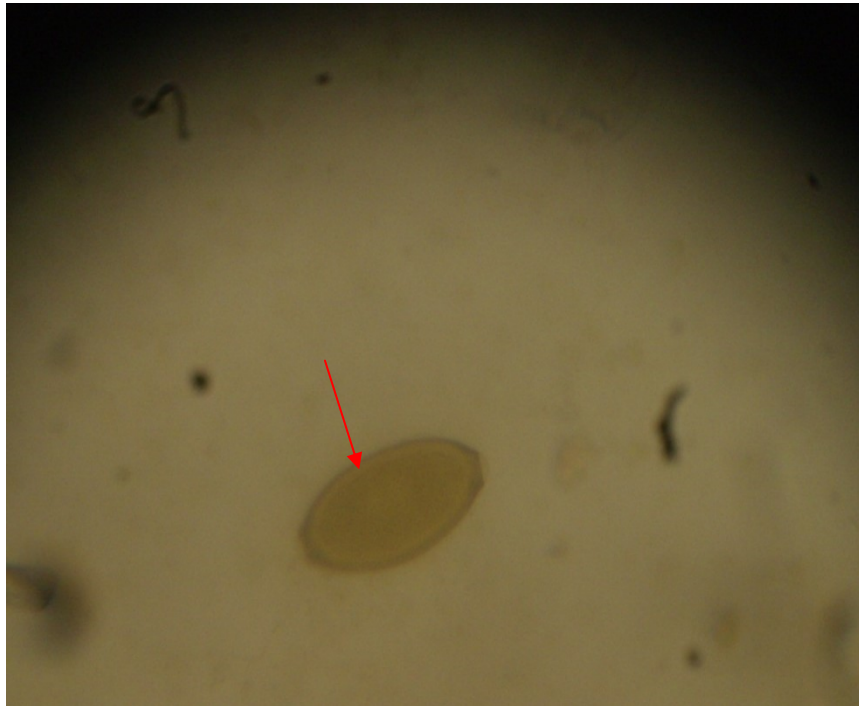


Plate 3: Image of *Trichuris trichiura* egg recovered from goat faeces

IV. DISCUSSION

This investigation revealed three helminth parasites comprising two nematodes, *Heamonchus contortus* and *Trichuris trichiura* and one cestode, *Taenia* species. An overall prevalence of 77.6% of endoparasite helminthes is reported in this present study. This result is similar to those obtained by [35] 75.5% in Port Hartcourt, [11] 75.75% in Ibadan and [36] 74.00% in Bangladesh. However, this finding is lower compared to 88.1%, 85.22%, and 89.33% reported by [37] in South West China, [38] in India, and [39] in Egypt respectively. The most prevalent parasite encountered was *H. contortus* (49.4%) which contrast the result of [15], who reported *S. papillosus* as the most prevalent helminth parasite in Nsukka urban, Nigeria. However, this result is not in conformity with the findings of [40] in Rwanda, who reported 71.8% prevalence of *H. contortus* in goats. There was higher endoparasitic infection in Red Sokoto goats (78.35%) than in West Africa Dwarf goats (73.2%). This difference in infection between breeds could be explained through the grazing habit of potential hosts. Some investigators opined that susceptibility to infection could be as a result of some breeds grazing close to the ground where their droppings are, therefore picking up eggs/oocysts or larvae [41]. The highest worm burden and prevalence rate of 43.6% and 51.7% was recorded for *H. contortus* in the two breed of goats. This high worm burden could be explained by the fact that immune compromised animals usually have high fecundity of parasites, due to malnutrition as shown by the investigated goats. Therefore, this makes it impossible for these animals to resist invasion of parasites [42, 43].

As regard sex, female goats were found to be more infected than their male counterparts, which is consistent with the reports of [3, 15, 44, 45, 46] who gave similar report between sex in goats. It should be noted that when female goats are pregnant or lactating, they experience hormonal changes which lowers their immunity and resistance to parasitic invasion, thus having high worm burden [3].

As regard age, adult goats above 2 years were more susceptible to infection than the young ones < 2 years. This finding is in consonance with the work of [10], who reported age as a risk factor for more susceptibility of infection in older small ruminant > 2years. Similarly, [47] disclosed that small ruminant adult goats have higher worm burden without adverse effect which leads to chronic infection.

In this present study, it was found that months and seasons have effect on endoparasite infection of small adult ruminants, throughout the period of investigation. It was found that *H. contortus* worm burden increased from July to September where it formed a single peak, then declined until December. This finding is consistent with the work of [47] who reported that adult *H. contortus* worm burden was high from May to October in Guinea Savannah zone of Nigeria. However, *T. trichiura* prevalence in goats increased from July

to September where it formed a small peak of 22%, then declines in October and then rises to its maximum levels of 37% in December. This finding accedes with [48] in Srinagar District of Kashmir, who reported that *Trichuris* count in Ovines increased in Autumn (42.02%), reached it maximum levels in Winter (59.37) and then tended to decline until Spring (53.22%) and reached minimum levels in Summer (30.6%) before increasing again in mid-Autumn. This study reported the prevalence of 7.9% *Taenia* spp in the study area, which is lower than 15.81% documented by [49] in Nigeria. The presence of zoonotic helminths (*Taenia* spp) in food animals slaughtered for human consumption in this study is of Public health concern. This finding is in consonance with the report of [29] who reported that these parasites are associated with different conditions in humans, ranging from diarrhea, intellectual and cognitive retardation, and retarded growth. In the present study, the seasonal effects on gastrointestinal parasites of small ruminant goats, was significant in early rainy season (July to August), late rainy season (September to October) and early dry season (November to December). Seasonal prevalence was highest (42.2%) in late rainy season, followed by 36.3% in early rainy season and finally 22.4% in dry season. This finding is in agreement with the result of [46] who reported highest seasonal prevalence in the rainy season.

V. CONCLUSION

In conclusion, this study was conducted to determine the prevalence of endoparasites in two breed of goats, WAD and RS goats. Three helminth parasites *H. contortus*, *T. trichiura* and *Taenia* species were recovered from 532 faecal samples of slaughtered goats at three abattoirs in Calabar, through corproscopy. An overall endoparasite prevalence rate of 77.6% was found in goats in this study. Red Sokoto were more infected than West Africa Dwarf goats. It was therefore recommended that small ruminant goat farmers in the study area should concentrate in the breeding of WAD goats, which are more resistant to endoparasite helminth infection than the RS goats. To ensure food safety for consumers of small ruminants in the study area and beyond, quality veterinary meat inspection is suggested to curtail the transmission of these helminthes to humans.

ACKNOWLEDGEMENT

I sincerely thank the Senior technologist of Biology laboratory of Cross River University of Technology for her assistance in bench work and identification of the parasites.

CONFLICT OF INTEREST

The author declared no conflict of interest, as there was no financial support either by company or individual.

REFERENCES

- [1]. Lawal-Adebawale OA (2012). Dynamics of ruminant livestock management in the content of the Nigerian Agricultural system, Livestock production, Khalid Javel, hitch open, DOI: 10.5772/52923. Available from:

- <http://www.intechopen.com/books/livestock-production/dynamics-of-ruminant-livestock-management-in-the-content-of-the-nigerian-agricultural-system>.
- [2]. Anyaegbunam IC, Okafor OJ (2013). Tripanosomiasis in Red Sokoto and West Africa Dwarf goats in Ikpa abattoirs, Nsukka Enugu State, Nigeria. *J Entomol Zool Stud* 1: 35-37.
 - [3]. Ma'azu A, Aliyu IW, Mao PS, Danladi SI (2018). Gastrointestinal parasites of goats in a Semi-Arid zone of Nigeria: Its prevalence and economic significance. *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)* 11(3): 37-41.
 - [4]. Fabusoro E, Lawal-Adebawale OA, Akinloye AK (2007). A study of rural livestock farmers' patronage of veterinary sciences for healthcare of small farm animals in Ogun State. *Nigerian Journal of Animal Production*, 34(1): 132-138.
 - [5]. Otchere EO (1986). Traditional cattle production in the sub-humid zone of Nigeria. In Von Kaufmann, R, Chater, S, and Blench, R., eds. *Livestock Research in Nigeria Sub-humid zone*. Proceedings of the second ILCA/NAPRI symposium held in Kaduna, Nigeria, 29 October- 2 November 1984.
 - [6]. Hirst KK (2017). The domestication history of goats (*Capra hircus*). Available on at: <https://www.throughco.com/the-domestication-history-of-goat-170661>. Accessed 2020.
 - [7]. FAOSTAT (2014). Data/Liver Animal/Goats. Data Available at: <https://www.fao.org/faostat/en/#data/PA>.
 - [8]. Yusufu OA, Kassali R, Aremu FJ, Ojo MO (2017). Market analysis of smallholder goat enterprise under tropical conditions. *Agricultura Tropica ET Subtropica*, 15(3): 121-127.
 - [9]. Dicerbo AR, Manfredi M, Zanzani S, Stradiotto K (2010). Gastrointestinal infection in goat farm in Lombardy (Northern Italy): analysis on community and spatial distribution of parasites. *Small Rum Res*. 88: 102-112.
 - [10]. Anugrah, SV Singh, Singh JP, Ramakant, Singh NK, Varum VK (2018). Epidemiology of gastrointestinal parasites in goats of Kumarganj region of Uttar Pradesh. *Journal of Pharmacognosy and Phytochemistry*. SP 4: 16-20.
 - [11]. Olanike AO, Olayide AJ, Oludunsin FO, Racheal AO, Japhet Dw (2015). Prevalence of gastrointestinal parasites of goats in Ibadan, Southwest, Nigeria. *World Journal of Agricultural Research*. 3(2): 49-51.
 - [12]. Adamu B, SamailaandBalarabe, Musa L (2006). Prevalence of haemoparasites of sheep and goats slaughtered in Bauchi abattoir *IJABR* 4(1&2): 128-133
 - [13]. Nwosu CO, Madu PP, Richards WS (2007). Prevalence and seasonal changes in the population of gastrointestinal nematodes of small ruminants in the semi-arial zone of north eastern Nigeria. *Vet Parasitol*. 144: 118-124
 - [14]. Maingi N, Gichanga EJ, Gichovi VM (1993). Prevalence of gastro-intestinal helminthes and coccidial parasites and frequency distribution of some nematode genera in some farms in four distribution of some nematode genera in some farms in four districts in Kenya. *Bull. Anim. Health prod. Afr*. 41: 285-290
 - [15]. Okoye IC, Obiezue RN, Okoye DN, Awi M (2013). High prevalence of gastro-intestinal parasites in indigeneous goats of Nigeria. *Paripex-Indian Journal of Research* 2(10): 17- 19.
 - [16]. Adedipe NO, Bakshi JS, Odegbare OA, Aliyu A (1996). Evolving the Nigeria Agricultural Research Strategy plan. Agro-ecological impacts, National Agricultural Research Project (NARP), 1996.
 - [17]. Urquhart GM, Armour J, Duncan JL, Jennings FW (2003). *Veterinary Parasitology*. Second edition, Blackwell Publishers, pp. 157-158.
 - [18]. Torres-Acosta J, Hoste H (2008). Alternative or improved methods to limit gastrointestinal parasitism in grazing sheep and goats. *Small ruminant Research*. 77(2): 48-59.
 - [19]. Calvete C, Ferrer L, Lacosta D, Calavia R, Ramos J, Ruiz-de-Arkaute M, Uriarte J (2014). Variability of the egg hatch assay to survey Benzimidazole resistance in nematode of small ruminants under field conditions. *Vet Parasitol*. 203(1): 102-113.
 - [20]. Bishop S, Morris C (2007). Genetics of disease resistance in sheep and goats. *Small ruminant research* 70(1): 48-59.
 - [21]. Odeniran PO, Jegede HO, Adewoga TOS (2016). Prevalence and risk perception of adult-stage parasites in slaughtered food animals (Cattle, sheep and goat) among local meat personel in Ipata abattoir, Iloriusn Nigeria. *Vet Med Anim Sci*. 4(1): 1.
 - [22]. Rabiun BM, Jegede OC (2010). Incidence of bovine cysticercus in Kano State, North- Western, Nigeria. *Bayero J Pure Appl Sci*. 3(1): 100-3.
 - [23]. Okolugbo BC, Luka S, Ndams IS (2014). Enzyme-linked immunosorbent Assay in the serodiagnosis of hydatidosis in camels (Camel dromedaries) and cattle in Sokoto, northern Nigeria. *Int J Infect Dis*. 13(1): 1-6
 - [24]. Adedipe OD, Uwalaka EC, Akinseye VO, Adediran OA, Cadmus SIB (2014). Gastrointestinal helminthes in slaughtered cattle in Ibadan, South-Western Nigeria. *J Vet Med*. 2014: 923561.
 - [25]. Ani OC, Nshiwu GN (2015). Assessment of intestinal parasites in goats slaughtered at Abakaliki abattoir, Ebonyi State, Nigeria. *Nig J Parasitol* 36(2): 81-4.
 - [26]. Nwoke EU, Odiko-mnoro OO, Ibiam GA, Umah OV, Ariom OTN (2015). Survey of common helminthes of goats slaughtered at Ankpa abattoir, Kogi State, Nigeria. *J Parasitol Vect Biol*. 7(5): 89-93.
 - [27]. Karshima NS, Bata SI, Bobbo AA (2016a). Prevalence risk factors and economic losses associated with fasciolosis in slaughtered cattle in Bauchi, North-Eastern Nigeria. *Alex J Vet Sci* 50(1): 87-93.
 - [28]. Magaji AA, Oboegbule SI, Daneji AI, HS, Salhu MD, Junaidu AU, et al. (2011). Incidence of hydatid cyst disease in food animals slaughtered at Sokoto Central abattoir, Sokoto State, Nigeria. *Vet Wld*. 4(5): 197-200.
 - [29]. Hotez PJ, Brooker S, Bethony JM, Bottazzi ME, Loukas A, Xiao S (2004). Current concepts: hookworm infection. *N Engl J Med* 351: 799-807.
 - [30]. Diop AG, de-Boer HM, Mandlhate C, Prilipko L, Meinardi H (2003). The global campaign against epilepsy in Africa. *Act Trop*. 87: 149-59.
 - [31]. Holst PJ and Denney (2003). How to tell the age of goat. *Agfact A7.2.2*. New South Wales.
 - [32]. Cheesbrough M (1987). A modification of the formol ether concentration technique for increased sensitivity in detection of *Schistosoma mansoni* eggs. *Amer. J. Trop. Med. Hyg*. 25: 87-97.
 - [33]. Whitlock HV (1948). Some modification of the McMaster helminth egg-counting technique and apparatus. *J. council Scient. Ind. Res*. 21: 177-180.
 - [34]. Soulsby EJL (1982). *Helminths, Arthropods and Protozoa of domesticated animals*. 7th Edn. Lea and Febiger, London, UK, ISBN: 9780812107807, pages 809
 - [35]. Owhoeli O, Elole K, Gboeloh (2014). Prevalence of gastrointestinal helminthes in exotic and indigenous goats slaughtered in selected abattoirs in Port Harcourt, South-South, Nigeria. Volume 2014/Article ID435913/8 pages/<https://doi.org/10.1155/2014/435913>
 - [36]. Nuruzzaman M, Haque MH, Sarker S, Begun N (2012). Abomasal nematodes in goats slaughtered at different abattoir of Thakurgaon District, Bangladesh. *Journal of Scientific Research* 4(2): 491-497.
 - [37]. Hao G, He X, Rui XU (2018). Prevalence of gastrointestinal parasites in Black goats in Liangshan Prefecture, SouthWest China. *Asian Journal of Animal and Veterinary Advances* 13: 346-351.
 - [38]. Pathak AK, Pal S (2008). Seasonal prevalence of gastrointestinal parasites in goats from Durg District of Chhattisgarh, India *Vet World*. 1(5): 136-137
 - [39]. Hassan NMF, Farag TK, Abu El Ezz, MNT, Abou-Zeina HAA (2019). Prevalence assessment of gastrointestinal parasitic

- infections among goats in Giza Governorate, Egypt. Bulletin of the National Research Centre. 43, 127 (2019) <http://doi.org/10.1186/s42269-019-0151-5>
- [40]. Mushonga B, Habumugisha D, Kandiwa E, Madzingira O, Samkange A, Segwagwe BE, Jaja IF(2018). Prevalence of Haemonchus contortus infections in sheep and goats in Nyagatare District, Rwanda. Journal of Veterinary Medicine Vol. 2018: Article ID 3602081, 9 pages. <https://doi.org/10.1155/2018/3602081>.
- [41]. Allonby EW (1980). Strategy and cost of helminth control with indications for research In: Impact of animal disease research and control on livestock production in Africa. Proceedings of the conference held in Nairobi Sept. 1980 OAU/ILADILCA/DSE, Berlin. 1980; pp. 135-140
- [42]. [Etter E, Chartier C, Hoste H, Borgida LP (1999). The influence of nutrition on the Peri parturient rise in faecal egg counts in dairy goat. Result from a two years study. Rev. de Med. 12: 975-980
- [43]. Emery DL, Hunt PW, Le jambre LF (2016). Haemonchus contortus : the then and now, and where to from here. International Journal for Parasitology. 46(12): 755-769.
- [44]. Valcarcel F, Carcia RC (1999). Prevalence and seasonal pattern of Caprine trichostrongyles in a dry area of central Spain. J Vet Med 6: 673-680
- [45]. Dagnachew S, Amanute A, Temesgen W. (2011). Epidemiology of gastrointestinal helminthiasis of small ruminants in selected sites of North Gondar Zone, Northwest Ethiopia. Ethiop Vet J 15(2): 57-68
- [46]. Rahman Md A, Labony SS, Dey AR, Alam MZ (2017). An epidemiological investigation of gastrointestinal parasites of small ruminants in Tangail, Bangladesh. Journal of BangladeshAgricultural University (2): 255-259
- [47]. Gana JJ, Makun H, Chiezey NP, Tekdek LB (2015). Epidemiological study on abomasal nematodes in slaughtered small ruminants raised in the Guinea Savannah zone of Nigeria. Sokoto Journal of Veterinary Sciences 13(2): 26-33.
- [48]. Gul N, Tak H (2016). Prevalence of Trichuris in small ruminants slaughtered in Srinagar District (T&K). J Parasit Dis 40(3): 741-744.
- [49]. Karshima SN, Maikai B-V, Kwaga JKP (2018). Helminths of veterinary and zoonotic importance in Nigerian ruminants: a 46 year meta-analysis (1970-2016) of their prevalence and distribution. Infectious Diseases of Poverty 7(52): 2018.